



## Nitrate removal by *Thiobacillus denitrificans* immobilized on poly(vinyl alcohol) carriers

Zhenya Zhang<sup>a</sup>, Zhongfang Lei<sup>b,\*</sup>, Xiaoyan He<sup>a</sup>, Zhiyin Zhang<sup>b</sup>, Yingnan Yang<sup>a</sup>, Norio Sugiura<sup>a</sup>

<sup>a</sup> Graduate School of Life and Environmental Sciences, University of Tsukuba, Tsukuba 305-8572, Japan

<sup>b</sup> Department of Environmental Science and Engineering, Fudan University, 220 Handan Road, Shanghai 200433, China

### ARTICLE INFO

#### Article history:

Received 8 March 2008

Received in revised form 1 July 2008

Accepted 16 July 2008

Available online 23 July 2008

#### Keywords:

Polyvinyl alcohol (PVA)

Boric acid

Freezing/thawing

*Thiobacillus denitrificans*

Denitrification

### ABSTRACT

Nitrate contamination is becoming a widespread environmental problem, and autotrophic denitrification with *Thiobacillus denitrificans* is a promising process considering efficiency, cost and maintenance. The denitrification efficiencies of *T. denitrificans* were compared in batch reactors between free cells and cells immobilized on polyvinyl alcohol (PVA) carriers made with thrice freezing/thawing and boric acid methods. The results indicated that the free cell reactor of *T. denitrificans* added with 10% (v/v) of PVA carrier made by thrice freezing/thawing (PVA-TFT) exhibited faster in  $S_2O_3^{2-}$ -S consumption,  $SO_4^{2-}$  generation, and  $NO_3^-$ -N denitrification, with corresponding values being 165 mg ( $S_2O_3^{2-}$ -S)/L d, 491 mg ( $SO_4^{2-}$ )/L d, and 44 mg ( $NO_3^-$ -N)/L d, which were increased by 50%, 61%, and 57% respectively compared to the control reactor with only free cells. Inhibition of denitrification by accumulated  $SO_4^{2-}$  in PVA-TFT reactor appeared at the concentration of approximately 6000 mg ( $SO_4^{2-}$ )/L, and 75% of  $NO_3^-$ -N removal efficiency was achieved after 12 d operation under the condition of initial 700 mg/L  $NO_3^-$ -N concentration.

© 2008 Elsevier B.V. All rights reserved.

### 1. Introduction

Polyvinyl alcohol (PVA), a cheap and synthetic polymer, has been widely used now for cell and enzyme immobilizations mainly because of its high durability, high chemical stability and non-toxicity to microorganisms or enzymes. Among the reported PVA immobilization methods, PVA-boric acid and freezing/thawing methods are frequently used.

Although two main problems, e.g., agglomeration of PVA gel beads and toxicity of saturated boric acid, cannot be absolutely resolved, PVA-boric acid method is still adopted for immobilizations of activated sludge [1–6], some functional microorganisms [5,7–11], and enzymes [12,13]. To overcome these problems, several modifications have been found effectively to enhance the properties of formed PVA beads, such as adding alginate [3,10,11] or powdered activated carbon [5], being treated with sodium sulfate [13] or by phosphorylation [2–4], or using glutaraldehyde to reduce the hydration [9], etc. Meanwhile, PVA, a kind of hot water soluble polymer, can be transformed to a stable, macroporous hydrogel under low temperature, which forms the basis of freezing/thawing method. Because of no requirement of toxic boric acid and increased mechanical strength of formed beads,

PVA-freezing/thawing method has been successfully used in cell immobilizations [14–19].

It has been noted that PVA beads made by these two methods exhibited porous structures from surface to interior [9,15,17,20], which is beneficial for adsorption of microorganisms and protection against detrimental conditions when used as carriers. However, little information can be found in literature about the application of porous PVA carrier directly in water or wastewater treatment.

On the other hand, nitrate contamination, originated from agricultural runoff, landfill leachate, leaking septic tanks, etc., is becoming a widespread environmental problem. And biological denitrification processes, including autotrophic denitrification and heterotrophic denitrification, have been commonly used for wastewater treatment. Compared with heterotrophic denitrification, autotrophic denitrification is attracting increasing interest in recent years due to the following three major advantages: (i) no residual organic problems because of the utilization of inorganic substances as electron donors; (ii) low operation and maintenance costs due to no external organic carbon needed; and (iii) lower cell yield, which minimizes sludge handling or lessens the risk of biological regrowth in distribution systems and disinfection by-product formation [21–27].

Among the denitrifying microorganisms, only a few species of autotrophic bacteria can reduce nitrate to nitrogen gas while oxidizing elemental sulfur or reduced sulfur compounds ( $S^{2-}$ ,  $S_2O_3^{2-}$ ,  $SO_3^{2-}$ ) to sulfate, and *T. denitrificans* is the most frequently used.

\* Corresponding author. Tel.: +86 21 65642018; fax: +86 21 65642018.  
E-mail address: [zfllei@fudan.edu.cn](mailto:zfllei@fudan.edu.cn) (Z. Lei).

Up to now, many researchers have paid more attention to the efficiencies and influence factors of autotrophic denitrification processes by *T. denitrificans* (enriched sludge or pure culture) with reduced sulfur compounds as electron donors in the treatment of nitrate-contaminated drinking water, groundwater or wastewaters [23–35]. Few of them demonstrated the efficiency of immobilized *T. denitrificans*, which might enhance the denitrification efficiency and protect the bacteria against detrimental environments.

The objective of this study was to investigate the denitrification efficiencies of *T. denitrificans* immobilized on PVA carriers formed by PVA–boric acid method and PVA–freezing/thawing method under the condition of thiosulfate used as reduced sulfur compound. And whether the immobilization processes could enhance the reactor performance or not compared with free cells was also discussed.

## 2. Materials and methods

### 2.1. Microorganisms and culture medium

*T. denitrificans* 3870 used in this study was obtained from the Institute of Physical and Chemical Research, Japan (RIKEN), which was separated from residual sludge of a domestic wastewater treatment plant.

The basal mineral medium utilized in this study contained (per liter): 1.8 g  $\text{KH}_2\text{PO}_4$ , 1.2 g  $\text{Na}_2\text{HPO}_4$ , 0.1 g  $(\text{NH}_4)_2\text{SO}_4$ , 0.1 g  $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ , 30 mg  $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ , 5.0 g  $\text{KNO}_3$ , 0.5 g  $\text{NaHCO}_3$ , and 100 ml of 10% (10 g/100 ml)  $\text{Na}_2\text{S}_2\text{O}_3 \cdot 5\text{H}_2\text{O}$ , which was suggested by RIKEN. Before use, the basal mineral medium (except  $\text{Na}_2\text{S}_2\text{O}_3$  solution) was autoclaved at 125 °C for 15 min and the  $\text{Na}_2\text{S}_2\text{O}_3$  solution was sterilized by filtration.

The culture medium was composed of 90% (v/v) of basal mineral medium and 10% (v/v) of trace element solution. The latter solution contained (mg/L): EDTA (500.0),  $\text{CaCl}_2$  (55.4),  $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$  (15.7),  $\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$  (16.1),  $\text{MnCl}_2 \cdot 4\text{H}_2\text{O}$  (50.6),  $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$  (220.0),  $(\text{NH}_4)_6\text{Mo}_7\text{O}_{24} \cdot \text{H}_2\text{O}$  (11.0), and  $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$  (49.9) [36].

All the chemicals used in the experiments were of chemical grade.

### 2.2. Experiment setup and efficiency assessment

All batch experiments were started at initial culture pH 7.0 (adjusted with 0.1 M NaOH) and conducted on a shaking table at  $30 \pm 2$  °C in duplicate in 500 ml reactors sealed with butyl rubber

stoppers. The working volume was 440 ml, composed of 400 ml of culture medium and 40 ml of the 4th-day *T. denitrificans* enrichment with the same culture medium. Before operation, the reactors were flushed with nitrogen gas for 3 min to exclude oxygen, and then all the outlets of the reactors were immersed in water.

The initial cell densities, indicated by optical density at 650 nm, were identical for all the reactors of the following experiments before startup ( $\text{OD}_{650} = 0.197$ ). And the initial  $\text{S}_2\text{O}_3^{2-}\text{-S}$  and  $\text{NO}_3^- \text{-N}$  concentrations for the reactors were approximately 3850 mg ( $\text{S}_2\text{O}_3^{2-}\text{-S}$ )/L and 700 mg ( $\text{NO}_3^- \text{-N}$ )/L, respectively.

There are four kinds of reactors in this study. One is the control, with only free cells and no carrier added in the culture medium, the other three were labeled as PVA–TFT, PVA–BA1, and PVA–BA2 for the reactors with carriers added and prepared with the methods of PVA–thrice freezing/thawing, PVA–boric acid with water-washing, and PVA–boric acid without water-washing, respectively. And the volume ratios of added carrier to total volume (carrier fill) were approximately 10% for the three cell-immobilized reactors.

The four reactors were compared and assessed with  $\text{SO}_4^{2-}$  generation rate,  $\text{NO}_3^- \text{-N}$  denitrification rate, and  $\text{S}_2\text{O}_3^{2-}\text{-S}$  consumption rate, respectively. These values at  $j$ th d were calculated as follows:

$$\text{SO}_4^{2-} \text{ generation rate (mg/L d)} = \frac{[M]_j - [M]_i}{j - i} \quad (1)$$

$$\text{S}_2\text{O}_3^{2-} \text{ - S consumption rate or NO}_3^- \text{ - N denitrification rate (mg/L d)} = \frac{[M]_i - [M]_j}{j - i} \quad (2)$$

where  $[M]$  (mg/L) is the concentration of  $\text{SO}_4^{2-}$ ,  $\text{S}_2\text{O}_3^{2-}\text{-S}$  or  $\text{NO}_3^- \text{-N}$  in the reactor, and  $i$  (d) or  $j$  (d) is the operation time ( $j > i$ ,  $i \geq 0$ ). In addition,  $j = i + 2$  (d) ( $i = 0, 2, 4, \dots$ ) for the calculations of daily values, and  $j = 12$  or 18 (d) and  $i = 0$  (d) for average values in this study.

Besides,  $\text{NO}_3^- \text{-N}$  removal efficiency, calculated as Eq. (3), was also used.

$$\text{NO}_3^- \text{ - N removal efficiency (\%)} = \frac{[M]_0 - [M]_t}{[M]_0} \times 100 \quad (3)$$

where  $[M]_0$  and  $[M]_t$  are the initial and terminal  $\text{NO}_3^- \text{-N}$  concentrations in the reactor, respectively.

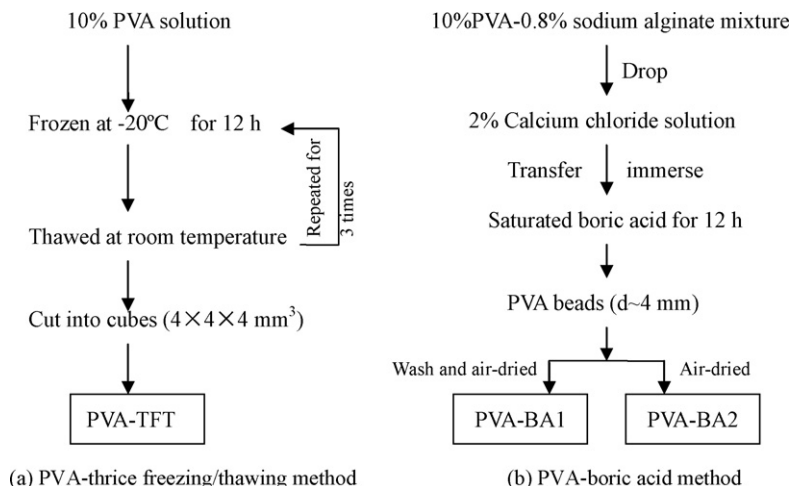


Fig. 1. Flowchart of carrier preparations with PVA–thrice freezing/thawing (PVA–TFT) (a) and PVA–boric acid (PVA–BA) (b) methods.

### 2.3. Preparation of PVA carriers

The PVA carriers were prepared following the procedures depicted in Fig. 1. PVA, with average polymerization degree of 2000, and sodium alginate were provided by Wako Pure Chemical Inc. Ltd., Japan.

The 10% (m/v) PVA solution was obtained by adding 10 g PVA to 100 ml of distilled water, which was heated to 60 °C to dissolve PVA.

In PVA–thrice freezing/thawing (PVA–TFT) method, the 10% PVA solution was first frozen at –20 °C for 12 h and later thawed at room temperature. After three cycles of freezing/thawing process, the resulting solidified PVA was cut into 4 mm × 4 mm × 4 mm cubes.

In PVA–boric acid (PVA–BA) method, the 10% PVA solution was kept at 4 °C for 24 h before 0.8% (m/v) sodium alginate being added. The mixture was extruded as drops into a 2% (m/v) calcium chloride solution, and later transferred into saturated boric acid and immersed for about 12 h to complete solidification. All these operations were carried out at room temperature under gently stirring, and the resulting beads were approximately 4 mm in diameter. As for PVA–BA1, the beads were completely washed with distilled water to remove any excess boric acid after solidification and added into reactor after air-dried; and for PVA–BA2, the beads were air-dried and added into reactor without water washing after solidification in boric acid.

### 2.4. Analytical methods

All the following indices were analyzed in duplicate, and the data present were the average values of duplicate experiments.

Biomass was indicated by optical density at the wavelength of 650 nm (OD<sub>650</sub>). OD<sub>650</sub> (DR4000U spectrophotometer, Hach Co.) and pH (Twin B-212, Horiba) were measured directly with sampled cultures.

For the determinations of nitrate nitrogen (NO<sub>3</sub><sup>–</sup>-N), sulfate (SO<sub>4</sub><sup>2–</sup>), and thiosulfate sulfur (S<sub>2</sub>O<sub>3</sub><sup>2–</sup>-S), the culture was sampled once every other day and centrifuged at 5000 rpm for 15 min, and the supernatant was used for the corresponding measurements.

The concentration of NO<sub>3</sub><sup>–</sup>-N was obtained with UV spectrophotometry (DR4000U, Hach Co.) by measuring the absorbance of supernatant at 220 nm and 275 nm, respectively [37]. SO<sub>4</sub><sup>2–</sup> was determined with Sulfaver 4 turbidimetric kit following the procedures developed by Hach Company [38]. In addition, a classical iodimetry [39] was used to quantify the amount of S<sub>2</sub>O<sub>3</sub><sup>2–</sup>-S remained in the culture.

### 2.5. Statistical analysis

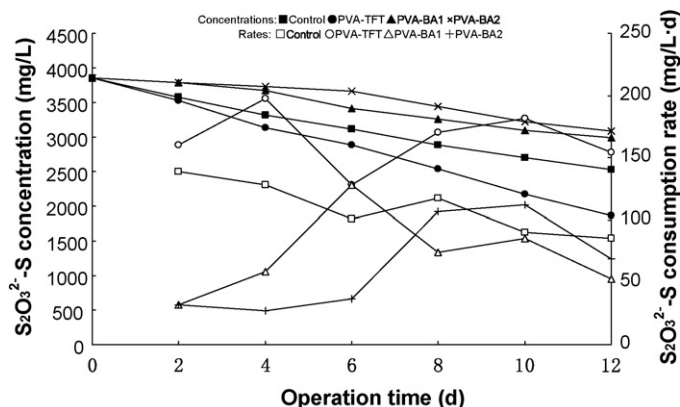
One-way ANOVA and *F*-test were used for statistical analysis, and *P* < 0.1, *P* < 0.05, *P* < 0.01, and *P* < 0.001 denoted significance levels of differences between cell-immobilized and control reactors.

**Table 1**

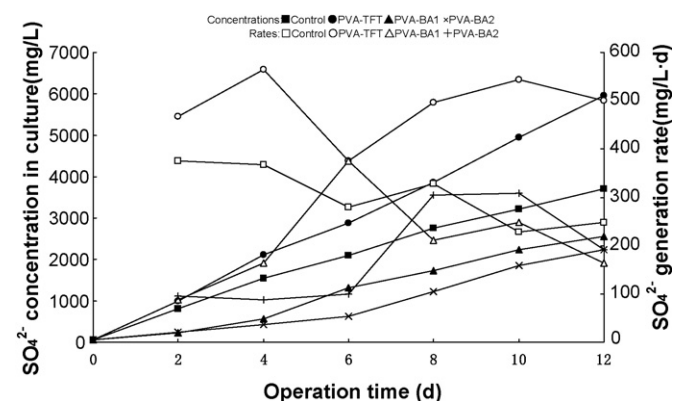
Results of S<sub>2</sub>O<sub>3</sub><sup>2–</sup>-S consumption, SO<sub>4</sub><sup>2–</sup> generation, and NO<sub>3</sub><sup>–</sup>-N denitrification rates obtained in the four reactors

Rates	Control	PVA–TFT	PVA–BA1	PVA–BA2
S <sub>2</sub> O <sub>3</sub> <sup>2–</sup> -S consumption rate (mg (S <sub>2</sub> O <sub>3</sub> <sup>2–</sup> -S)/Ld)	110.25 ± 21.37	165.33 ± 23.93 <sup>c</sup>	72.00 ± 33.03 <sup>b</sup>	64.00 ± 38.21 <sup>b, 0.71</sup>
SO <sub>4</sub> <sup>2–</sup> generation rate (mg (SO <sub>4</sub> <sup>2–</sup> )/Ld)	304.67 ± 62.20	491.33 ± 66.33 <sup>d</sup>	208.67 ± 98.09 <sup>a</sup>	181.33 ± 103.74 <sup>b, 0.65</sup>
NO <sub>3</sub> <sup>–</sup> -N denitrification rate (mg (NO <sub>3</sub> <sup>–</sup> -N)/Ld)	28.00 ± 6.20	44.00 ± 4.38 <sup>d</sup>	19.33 ± 9.61 <sup>a</sup>	15.33 ± 8.16 <sup>b, 0.46</sup>

Rate data expressed as mean ± S.D. Superscripts a, b, c, and d denote significance of differences between the cell-immobilized reactor and the control reactor at *P* < 0.1, *P* < 0.05, *P* < 0.01, and *P* < 0.001, respectively. And the numerical superscripts indicate the approximate significance levels of differences between PVA–BA1 and PVA–BA2 reactors obtained by ANOVA analysis.



**Fig. 2.** Variations of S<sub>2</sub>O<sub>3</sub><sup>2–</sup>-S concentrations and consumption rates in the four reactors.



**Fig. 3.** Variations of SO<sub>4</sub><sup>2–</sup> concentrations and generation rates in the four reactors. The SO<sub>4</sub><sup>2–</sup> concentration introduced by the addition of basal mineral medium and trace element solution has been excluded from the results.

## 3. Results and discussion

### 3.1. S<sub>2</sub>O<sub>3</sub><sup>2–</sup> consumption and SO<sub>4</sub><sup>2–</sup> generation

The variations of S<sub>2</sub>O<sub>3</sub><sup>2–</sup>-S and SO<sub>4</sub><sup>2–</sup> concentrations with the corresponding consumption or generation rates are illustrated in Figs. 2 and 3, respectively.

It can be seen that the S<sub>2</sub>O<sub>3</sub><sup>2–</sup>-S concentrations showed similar decreased trends in the four reactors, and the largest decreases were observed in the PVA–TFT reactor from the 4th day on (Fig. 2). The total decreases of S<sub>2</sub>O<sub>3</sub><sup>2–</sup>-S concentrations after 12 d operation were 1323, 1984, 864, and 768 mg (S<sub>2</sub>O<sub>3</sub><sup>2–</sup>-S)/L in the control, PVA–TFT, PVA–BA1, and PVA–BA2 reactors, respectively.

The variations of the S<sub>2</sub>O<sub>3</sub><sup>2–</sup>-S consumption rates seemed to be more complicated than S<sub>2</sub>O<sub>3</sub><sup>2–</sup>-S concentrations (Fig. 2). The *T. denitrificans* immobilized on PVA–BA carriers exhibited much lower activities, indicated by S<sub>2</sub>O<sub>3</sub><sup>2–</sup>-S consumption rate, than free cells and those immobilized on PVA–TFT carrier before the 6th day,

although complete washing in the making process of PVA beads might have some beneficial effect on eliminating the toxicity of boric acid (Fig. 2;  $P < 0.05$  for both PVA–BA reactors compared to the control, Table 1). In this study, the average  $S_2O_3^{2-}$ –S consumption rates were approximately 110, 165, 72, and 64 mg ( $S_2O_3^{2-}$ –S)/L/d for the control, PVA–TFT, PVA–BA1, and PVA–BA2 reactors, respectively (Table 1). And significant increase was observed for  $S_2O_3^{2-}$ –S consumption rate in the PVA–TFT reactor compared to the control ( $P < 0.01$ ), while no significant difference was found for  $S_2O_3^{2-}$ –S consumption between the two PVA–BA reactors ( $P = 0.71$ ) (Table 1).

The results of  $SO_4^{2-}$  concentrations and generation rates in the reactors were in accordance with those of  $S_2O_3^{2-}$ –S (Figs. 2 and 3). The total increases of  $SO_4^{2-}$  concentration were 3656, 5896, 2504, and 2176 mg/L, with average  $SO_4^{2-}$  generation rates approximately 305, 491, 209, and 181 mg/L/d for the control, PVA–TFT, PVA–BA1, and PVA–BA2 reactors, respectively (Table 1). Still, significant increase was observed for  $SO_4^{2-}$  generation rate in the PVA–TFT reactor compared to the control ( $P < 0.001$ ), and no significant difference was obtained between the two PVA–BA reactors ( $P = 0.65$ ) (Table 1). In addition, the  $SO_4^{2-}$  generation rate seemed to vary in the same way as  $S_2O_3^{2-}$ –S consumption rate in each reactor, resulting from the sulfur balance of the denitrification process.

From Figs. 2 and 3, *T. denitrificans* immobilized on PVA–TFT carrier seemed to use thiosulfate more efficiently than the others, including free cells. It was reported that PVA beads made by freezing/thawing with or without cell addition possessed distinctive heterogenous macropores [15,17,20,40]. The average inner dimensions of the macropores obtained by Szczesna-Antczak and Galas [17] and Hatakeyema et al. [40] were approximately 2–10  $\mu\text{m}$ , which is close to the dimension of microorganisms and beneficial for cell adsorption and immobilization. It was reported that microorganisms immobilized on porous carriers sintered with coal fly-ash could tolerate 15% of salinity and exhibited 99% of phenol removal in 16 h [41], suggesting that porous structure might provide protection for microorganisms from adverse conditions, e.g., high concentration of  $SO_4^{2-}$  in this study. Its detailed protection mechanism needs further investigations. Compared with the control reactor, the average  $S_2O_3^{2-}$ –S consumption and  $SO_4^{2-}$  generation rates in the PVA–TFT reactor were increased by 50% and 61%, respectively. Although the total  $SO_4^{2-}$  concentration increased to 5896 mg/L in the PVA–TFT reactor after operation for 12 d, no apparent inhibition was observed. The obtained  $SO_4^{2-}$  concentration, higher than the initial inhibition concentration (5000 mg/L) observed by Claus and Kutzner [22] for a pure culture of *T. denitrificans*, might be resulted from the protection effect of porous PVA structures. In addition, this  $SO_4^{2-}$  concentration is much higher than 1500 mg/L (500 mg  $SO_4^{2-}$ –S/L) [42] and 2000 mg/L [43] obtained in enriched mixed cultures of autotrophic organisms, indicating that *T. denitrificans* could tolerate higher sulfate concentration conditions when immobilized on porous materials.

Unlike in cell-immobilized reactors, the  $SO_4^{2-}$  generation rates in the control seemed to decrease with the increase of  $SO_4^{2-}$  concentration. However, that the inhibition caused by  $SO_4^{2-}$  accumulation started at which  $SO_4^{2-}$  level could not be obtained according to our experiments because of the complexity of denitrification process and multi-factors involved.

### 3.2. Denitrification efficiency

Seen from Fig. 4, the  $NO_3^-$ –N concentrations in the reactors decreased with time going on. The PVA–TFT reactor with *T. denitrificans* immobilized exhibited the highest  $NO_3^-$ –N removal among the four reactors, about 75% of  $NO_3^-$ –N being removed in all after 12 d operation under the same initial  $NO_3^-$ –N concentration (~700 mg ( $NO_3^-$ –N)/L). Compared with the control reactor

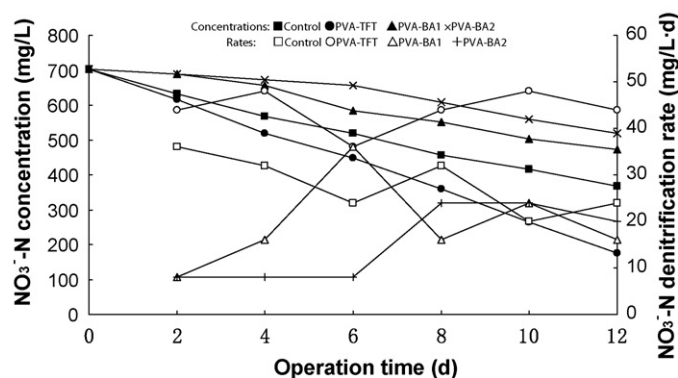


Fig. 4. Variations of  $NO_3^-$ –N concentrations and denitrification rates in the four reactors.

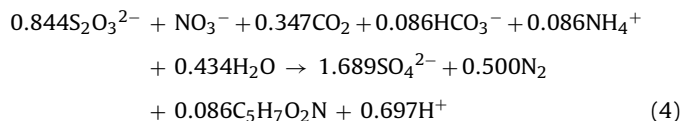
(free cells), in which 52% of  $NO_3^-$ –N removed after 12 d operation, the total  $NO_3^-$ –N removal efficiency of the PVA–TFT reactor was increased by 27%.

The total decreases of  $NO_3^-$ –N concentration were 336, 528, 232, and 184 mg/L, and the average  $NO_3^-$ –N denitrification rates were 28, 44, 19, and 15 mg ( $NO_3^-$ –N)/L/d, respectively, for the control, PVA–TFT, PVA–BA1, and PVA–BA2 reactors (Table 1). Compared with free cells, *T. denitrificans* exhibited higher denitrification activity on PVA–TFT carrier, with the average  $NO_3^-$ –N denitrification rate increased by 57%. A significant increase was also observed for  $NO_3^-$ –N denitrification rate in the PVA–TFT reactor in comparison with the control ( $P < 0.001$ ), and no significant difference was observed between the two PVA–BA reactors ( $P = 0.46$ ) (Table 1). The average denitrification activities for those cells immobilized on PVA–BA carriers decreased ( $P < 0.1$  for PVA–BA1 and  $P < 0.05$  for PVA–BA2, respectively) (Table 1), although some recovery from the toxicity of boric acid was observed after operation for 6–8 d (Fig. 4). The variations of  $NO_3^-$ –N in the four reactors were in agreement with the results of  $S_2O_3^{2-}$ –S and  $SO_4^{2-}$  obtained in this study.

In addition, the terminal pH values after 12 d operation were 6.53, 6.31, 6.66, and 6.78 in the control, PVA–TFT, PVA–BA1, and PVA–BA2 reactors, respectively, under the same initial pH of 7.0, which lay between 6 and 9, the optimum pH range for *T. denitrificans* [44].

### 3.3. Stoichiometric analysis

Eq. (4) can be used to represent the stoichiometric reaction when thiosulfate being used as reduced sulfur compound in denitrification process [42,45]:



According to the above reaction, the theoretical stoichiometric values of  $\Delta S_2O_3^{2-}$ –S/ $\Delta NO_3^-$ –N,  $\Delta SO_4^{2-}$ / $\Delta NO_3^-$ –N and  $\Delta SO_4^{2-}$ / $\Delta S_2O_3^{2-}$ –S ratios were calculated as 3.86 g/g, 11.58 g/g, and 3.00 g/g, respectively, which were quite similar to the results obtained in this study (Table 2).

It can be seen that all the relative errors were less than  $\pm 10\%$ . The relative errors for  $SO_4^{2-}$  quantifications with turbidimetric method and  $NO_3^-$ –N determinations with UV spectrophotometry were pointed out to be 0.2–7.0% and 2.6–5.1%, respectively [37]. Therefore, taking the errors possibly brought by sampling and the analytical methods into consideration, the obtained results suggested that the denitrification processes completed by

**Table 2**  
Average ratios for  $\Delta S_2O_3^{2-}\text{-S}/\Delta NO_3^- \text{-N}$ ,  $\Delta SO_4^{2-}/\Delta NO_3^- \text{-N}$ , and  $\Delta SO_4^{2-}/\Delta S_2O_3^{2-}\text{-S}$  obtained in the four reactors

Ratios	Values (g/g)				Relative errors (%) <sup>a</sup>			
	Control	PVA-TFT	PVA-BA1	PVA-BA2	Control	PVA-TFT	PVA-BA1	PVA-BA2
$\Delta S_2O_3^{2-}\text{-S}/\Delta NO_3^- \text{-N}$	3.94	3.75	3.72	4.17	2.07	-2.85	-3.63	8.03
$\Delta SO_4^{2-}/\Delta NO_3^- \text{-N}$	10.88	11.17	10.79	11.83	-6.04	-3.54	-6.82	2.11
$\Delta SO_4^{2-}/\Delta S_2O_3^{2-}\text{-S}$	2.76	2.97	2.90	2.83	-8.00	-1.00	-3.33	-5.67

<sup>a</sup> The relative errors (RE) were calculated according to the following equation:  $RE(\%) = (V_d - V_t)/V_t \times 100$ , in which  $V_d$  and  $V_t$  were the determined and theoretical values, respectively.

*T. denitrificans* were progressing thoroughly and other nitrogenous by-products might not generated in the reactors.

### 3.4. Efficiency of the PVA-TFT reactor after prolonged-time operation

The above results showed the PVA-TFT reactor performed more efficiently than others in the denitrification process and 75% of 700 mg/L  $NO_3^- \text{-N}$  could be removed after 12 d operation. To assess the denitrification potential of this kind of immobilized *T. denitrificans* and the effect of generated  $SO_4^{2-}$  concentration on denitrification, a prolonged-time operation was carried out in another PVA-TFT reactor and all the related indices were analyzed and determined as above (Fig. 5).

The results indicated that the persistent increase trend of  $SO_4^{2-}$  concentration slowed down and the  $SO_4^{2-}$  generation rate began to decrease largely from the 12th day on, which was in agreement with the decrease trends of  $NO_3^- \text{-N}$  concentration and  $NO_3^- \text{-N}$  denitrification rate (Fig. 5). The average daily  $SO_4^{2-}$  generated and  $NO_3^- \text{-N}$  removed were about 490 mg ( $SO_4^{2-}$ )/Ld and 40 mg ( $NO_3^- \text{-N}$ )/Ld respectively before the 12th day, which were approximately 5.5 times of the corresponding daily values obtained during the period

of the 12th to 18th days. Moreover, the terminal pH of the reactor after 18 d operation was 5.8, which lay out of 6–9, the optimum pH range for *T. denitrificans* [44]. Thus, we concluded that  $SO_4^{2-}$  started to inhibit the denitrification of *T. denitrificans* at approximately 6000 mg/L, which is much higher than the findings of Campos et al. (1500 mg ( $SO_4^{2-}$ )/L or 500 mg ( $SO_4^{2-}\text{-S}$ )/L) [42] and Oh et al. (2000 mg ( $SO_4^{2-}$ )/L) [43] in mixed cultures of enriched autotrophic organisms, and also higher than 5000 mg ( $SO_4^{2-}$ )/L obtained by Claus and Kutzner [22] in a pure culture of *T. denitrificans*. The enhancement of sulfate tolerance could be attributed to the protection of porous PVA structures for immobilized *T. denitrificans*.

## 4. Conclusions

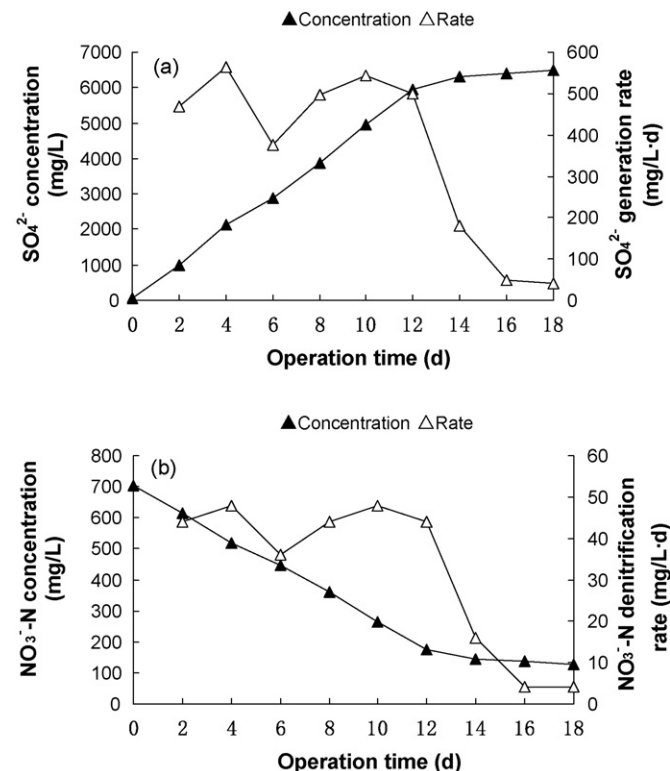
The present study showed that thrice freezing/thawing method could be used for the making of PVA carrier to immobilize *T. denitrificans* in the treatment of  $NO_3^- \text{-N}$  contained wastewaters. The *T. denitrificans* immobilized on PVA-TFT carrier could remove 75% of  $NO_3^- \text{-N}$  after 12 d operation when 700 mg/L of initial  $NO_3^- \text{-N}$  concentration being applied, and the average  $NO_3^- \text{-N}$  denitrification rate was 44 mg ( $NO_3^- \text{-N}$ )/Ld. The results of prolonged operation of PVA-TFT reactor indicated that *T. denitrificans* exhibited excellent denitrification activities when generated  $SO_4^{2-}$  concentration less than 6000 mg/L, implying that PVA-TFT carriers might perform better in continuously operated reactors in flow-in and flow-out mode, provided that appropriate design (including wastewater characteristics and operation parameters) being guaranteed. To ensure this kind of PVA carrier being put into practice, detailed investigations, including the inner structures of the carrier, effect of porous structure on sulfate or nitrate diffusion, duration of the immobilized cells, and optimized design of the reactor, should be followed.

## Acknowledgement

The authors acknowledged the help and kindness of the Institute of Physical and Chemical Research, Japan (RIKEN) in this study.

## References

- [1] S. Hashimoto, K. Furakawa, Immobilization of activated sludge by PVA-boric acid method, *Biotechnol. Bioeng.* 30 (1987) 52–59.
- [2] K.C. Chen, Y.F. Lin, Immobilization of microorganism with phosphorylated polyvinyl alcohol (PVA) gel, *Enzyme Microb. Technol.* 16 (1994) 79–83.
- [3] K.C. Chen, S.J. Chen, J.Y. Houg, Improvement of gas permeability of denitrifying PVA gel beads, *Enzyme Microb. Technol.* 18 (1996) 502–506.
- [4] K.C. Chen, S.C. Lee, S.C. Chin, J.Y. Houg, Simultaneous carbonitrogen removal in wastewater using phosphorylated PVA immobilized microorganisms, *Enzyme Microb. Technol.* 23 (1996) 311–320.
- [5] I.S. Chang, C.I. Kim, B.U. Nam, The influence of poly-vinyl-alcohol (PVA) characteristics on the physical stability of encapsulated immobilization media for advanced wastewater treatment, *Process Biochem.* 40 (2005) 3050–3054.
- [6] L. Zhang, W. Wu, J. Wang, Immobilization of activated sludge using improved polyvinyl alcohol (PVA) gel, *J. Environ. Sci.* 19 (2007) 1293–1297.
- [7] K. Hanaki, S. Hirunmasuwan, T. Matsuo, Protection of methanogenic bacteria from low pH and toxic materials by immobilization using polyvinyl alcohol, *Water Res.* 28 (1994) 877–885.



**Fig. 5.**  $SO_4^{2-}$  (a) and  $NO_3^- \text{-N}$  (b) profiles in the PVA-TFT reactor during a prolonged operation.

- [8] J.K. Seo, I.H. Jung, M.R. Kim, B.J. Kim, S.W. Nam, S.K. Kim, Nitrification performance of nitrifiers immobilized in PVA (polyvinyl alcohol) for a marine recirculating aquarium system, *Aquacult. Eng.* 24 (2001) 181–194.
- [9] C. Jeon, J.Y. Park, Y.J. Yoo, Novel immobilization of alginate for heavy metal removal, *Biochem. Eng. J.* 11 (2002) 159–166.
- [10] Z. Long, Y. Huang, Z. Cai, W. Cong, F. Ouyang, Immobilization of *Acidithiobacillus ferrooxidans* by a PVA-boric method for ferrous sulphate oxidation, *Process Biochem.* 39 (2004) 2129–2133.
- [11] Y. Wang, X. Yang, H. Li, W. Tu, Immobilization of *Acidithiobacillus ferrooxidans* with complex of PVA and sodium alginate, *Polym. Degrad. Stabil.* 91 (2006) 2408–2414.
- [12] O. Ariga, T. Yamakawa, H. Fujimatsu, Y. Sano, Immobilization of  $\beta$ -galactosidase with polyvinyl alcohol, *J. Ferment. Bioeng.* 68 (1989) 293–295.
- [13] A. Idris, N.A.M. Zain, M.S. Suhaimi, Immobilization of Baker's yeast invertase in PVA-alginate matrix using innovative immobilization technique, *Process Biochem.* 43 (2008) 331–338.
- [14] O. Ariga, H. Takagi, H. Nishizawa, Y. Sano, Immobilization of microorganisms with PVA hardened by iterative freezing and thawing, *J. Ferment. Technol.* 65 (1987) 651–658.
- [15] V.I. Lozinsky, A.L. Zubov, E.F. Titova, Poly (vinyl alcohol) cryogels employed as matrices for cell immobilization. 2. Entrapped cells resemble porous fillers in their effects on the properties of PVA-cryogel carrier, *Enzyme Microb. Technol.* 20 (1997) 182–190.
- [16] W.M. Rostron, D.C. Stuckey, A.A. Young, Nitrification of high strength ammonia wastewaters: comparative study of immobilisation media, *Water Res.* 35 (2001) 1169–1178.
- [17] M. Szczesna-Antczak, E. Galas, *Bacillus subtilis* cells immobilised in PVA-cryogels, *Biomol. Eng.* 17 (2001) 55–63.
- [18] G. Cao, Q. Zhao, X. Sun, T. Zhang, Characterization of nitrifying and denitrifying bacteria coimmobilized in PVA and kinetics model of biological nitrogen removal by coimmobilized cells, *Enzyme Microb. Technol.* 30 (2002) 49–55.
- [19] Y. Wang, Y. Tian, B. Han, H. Zhao, J. Bi, B. Cai, Biodegradation of phenol by free and immobilized *Acinetobacter* sp. strain PD12, *J. Environ. Sci.* 19 (2007) 222–225.
- [20] F. Yokoyama, I. Masada, K. Shimamura, T. Ikawa, K. Monobe, Morphology and structure of highly elastic poly(vinyl alcohol) hydrogel prepared by repeated freezing and melting, *Colloid Polym. Sci.* 264 (1986) 595–601.
- [21] B. Batchelor, A.W. Lawrence, Autotrophic denitrification using elemental sulfur, *J. Water Pollut. Control Fed.* 50 (1978) 1986–2001.
- [22] G. Claus, H.J. Kutzner, Physiology and kinetics of autotrophic denitrification by *Thiobacillus denitrificans*, *Appl. Microbiol. Biotechnol.* 22 (1985) 283–288.
- [23] A. Koenig, L.H. Liu, Autotrophic denitrification of landfill leachate using elemental sulphur, *Water Sci. Technol.* 34 (1996) 469–476.
- [24] A. Koenig, L.H. Liu, Kinetic model of autotrophic denitrification in sulphur packed-bed reactors, *Water Res.* 35 (2001) 1969–1978.
- [25] T.C. Zhang, D.G. Lampe, Sulfur:Limestone autotrophic denitrification processes for treatment of nitrate-contaminated water: batch experiments, *Water Res.* 33 (1999) 599–608.
- [26] H. Wang, J. Qu, Combined biochemical and sulfur autotrophic denitrification for drinking water treatment, *Water Res.* 37 (2003) 3767–3775.
- [27] R. Sierra-Alvarez, R. Beristain-Cardoso, M. Salazar, J. Gomez, E. Razo-Flores, J.A. Field, Chemolithotrophic denitrification with elemental sulfur for groundwater treatment, *Water Res.* 41 (2007) 1251–1262.
- [28] J.M. Flere, T.C. Zhang, Nitrate removal with sulfur-limestone autotrophic denitrification processes, *J. Environ. Eng.* 125 (8) (1999) 721–729.
- [29] L. Kuai, W. Verstraete, Autotrophic denitrification with elemental sulphur in small-scale wastewater treatment facilities, *Environ. Technol.* 20 (1999) 201–209.
- [30] R. Yamamoto-Ikemoto, T. Komori, M. Nomura, T. Matsukami, Nitrogen removal from hydroponic culture wastewater by autotrophic denitrification using thiosulphate, *Water Sci. Technol.* 42 (2000) 369–376.
- [31] M.I.M. Soares, Denitrification of groundwater with elemental sulfur, *Water Res.* 36 (2002) 1392–1395.
- [32] S. Vidal, C. Rocha, H. Galvao, A comparison of organic and inorganic carbon controls over biological denitrification in aquaria, *Chemosphere* 48 (2002) 445–451.
- [33] H.S. Moon, K.H. Ahn, S. Lee, K. Nam, J.Y. Kim, Use of autotrophic sulfur-oxidizers to remove nitrate from bank filtrate in permeable reactive barrier system, *Environ. Pollut.* 129 (2004) 499–507.
- [34] H.S. Moon, S.W. Chang, K. Nam, J. Choe, J.Y. Kim, Effect of reactive media composition and co-contaminants on sulphur-based autotrophic denitrification, *Environ. Pollut.* 144 (2006) 802–807.
- [35] Z. Zhao, W. Qiu, A. Koenig, X. Fan, J.D. Gu, Nitrate removal from saline water using autotrophic denitrification by the bacterium *Thiobacillus denitrificans* MP-1, *Environ. Technol.* 25 (2004) 1201–1210.
- [36] S. Hashimoto, K. Furakawa, M. Shoiyama, Enrichment of sulfur denitrifying bacteria and its accumulation to elemental sulfur, *Jpn. J. Water Pollut. Res.* 12 (7) (1989) 431–440.
- [37] State Environmental Protection Administration of China (SEPA), Standard methods for the determination of water and wastewater, fourth ed., China Environmental Science Press, Beijing, 2002, pp. 164–165, pp. 266–268 (in Chinese).
- [38] Hach Co., *Water Analysis Handbook, Method 8051*, 2003.
- [39] N. Saito, Y. Yoshino, K. Saito, M. Fujimoto, K. Mizumachi (Eds.), *Practical Analytical Chemistry*, Shokabo Publishing Co. Ltd., Tokyo, 1975, pp. 175–177 (in Japanese).
- [40] T. Hatakeyama, J. Uno, C. Yamada, A. Kishi, H. Hakakeyama, Gel-sol transition of poly(vinyl alcohol) hydrogels formed by freezing and thawing, *Thermochim. Acta* 431 (2005) 144–148.
- [41] Y. Li, Z. Lei, Z. Zhang, N. Sugiura, Effects of nutrient addition on phenol biodegradation rate in biofilm reactors for hypersaline wastewater treatment, *Environ. Technol.* 27 (2006) 511–520.
- [42] J.L. Campos, S. Carvalho, R. Portela, A. Mosquera-Corral, R. Mendez, Kinetics of denitrification using sulphur compounds: effect of S/N ratio, endogenous and exogenous compounds, *Bioresour. Technol.* 99 (2008) 1293–1299.
- [43] S.E. Oh, K.S. Kim, H.C. Choi, J. Cho, I.S. Kim, Kinetics and physiological characteristics of autotrophic denitrification by denitrifying sulphur bacteria, *Water Sci. Technol.* 42 (2000) 59–68.
- [44] D.H. Bergey, J.G. Holt, *Bergey's Manual of Determinative Bacteriology*, 9th ed., Lippincott Williams & Wilkins, Baltimore, USA, 1994, pp. 433–438.
- [45] J.J. Bisogni Jr., C.T. Driscoll Jr., Denitrification using thiosulfate and sulfide, *J. Environ. Eng.* 103 (1977) 593–604.